

i-STAT EC4+ Cartridge



NAME

i-STAT EC4+ Cartridge – REF 03P81-25

INTENDED USE

The i-STAT EC4+ cartridge with the i-STAT 1 System is intended for use in the *in vitro* quantification of sodium, potassium, glucose and hematocrit in arterial, venous, or capillary whole blood.

Analyte	Intended Use
Sodium (Na)	Sodium measurements are used for monitoring electrolyte imbalances.
Potassium (K)	Potassium measurements are used in the diagnosis and monitoring of diseases and clinical conditions that manifest high and low potassium levels.
Glucose (Glu)	Glucose measurements are used in the diagnosis, monitoring, and treatment of carbohydrate metabolism disorders including, but not limited to, diabetes mellitus, neonatal hypoglycemia, idiopathic hypoglycemia, and pancreatic islet cell carcinoma.
Hematocrit (Hct)	Hematocrit measurements can aid in the determination and monitoring of normal or abnormal total red cell volume status including, but not limited to, conditions such as anemia, erythrocytosis, and blood loss related to trauma and surgery.

SUMMARY AND EXPLANATION/CLINICAL SIGNIFICANCE

Measured:

Sodium (Na)

Tests for sodium in the blood are important in the diagnosis and treatment of patients suffering from hypertension, renal failure or impairment, cardiac distress, disorientation, dehydration, nausea and diarrhea. Some causes of increased values for sodium include dehydration, diabetes insipidus, salt poisoning, skin losses, hyperaldosteronism and CNS disorders. Some causes for decreased values for sodium include dilutional hyponatremia (cirrhosis), depletion hyponatremia and syndrome of inappropriate ADH.

Potassium (K)

Tests for potassium in the blood are important in the diagnosis and treatment of patients suffering from hypertension, renal failure or impairment, cardiac distress, disorientation, dehydration, nausea and diarrhea. Some causes of increased values for potassium include renal glomerular disease, adrenocortical insufficiency, diabetic ketoacidosis (DKA), sepsis and *in vitro* hemolysis. Some causes of decreased values for potassium include renal tubular disease, hyperaldosteronism, treatment of DKA, hyperinsulinism, metabolic alkalosis and diuretic therapy.

Glucose (Glu)

Glucose is a primary energy source for the body and the only source of nutrients for brain tissue. Measurements for determination of blood glucose levels are important in the diagnosis and treatment of patients suffering from diabetes and hypoglycemia. Some causes for increased values of glucose include diabetes mellitus, pancreatitis, endocrine disorders (e.g., Cushing's syndrome), drugs (e.g., steroids, thyrotoxicosis), chronic renal failure, stress, or I.V. glucose infusion. Some causes of decreased values of glucose include insulinoma, adrenocortical insufficiency, hypopituitarism, massive liver disease, ethanol ingestion, reactive hypoglycemia, and glycogen storage disease.

Hematocrit (Hct)

Hematocrit is a measurement of the fractional volume of red blood cells. This is a key indicator of the body's state of hydration, anemia or severe blood loss, as well as the blood's ability to transport oxygen. A decreased hematocrit can be due to either overhydration, which increases the plasma volume, or a decrease in the number of red blood cells caused by anemias or blood loss. An increased hematocrit can be due to loss of fluids, such as in dehydration, diuretic therapy, and burns, or an increase in red blood cells, such as in cardiovascular and renal disorders, polycythemia vera, and impaired ventilation.

TEST PRINCIPLE

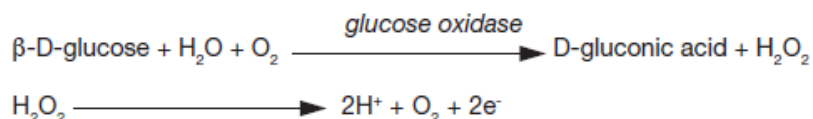
The i-STAT System uses direct (undiluted) electrochemical methods. Values obtained by direct methods may differ from those obtained by indirect (diluted) methods.¹

Measured:**Sodium (Na) and Potassium (K)**

The respective analyte is measured by ion-selective electrode potentiometry. Concentrations are calculated from the measured potential through the Nernst equation.

Glucose (Glu)

Glucose is measured amperometrically. Oxidation of glucose, catalyzed by the enzyme glucose oxidase, produces hydrogen peroxide (H₂O₂). The liberated H₂O₂ is oxidized at the electrode to produce a current proportional to the sample glucose concentration.

**Hematocrit (Hct)**

Hematocrit is determined conductometrically. The measured conductivity, after correction for electrolyte concentration, is inversely related to the hematocrit.

Calculated:

Hemoglobin (Hb)

The i-STAT System provides a calculated hemoglobin result which is determined as follows:

hemoglobin (g/dL) = hematocrit (% PCV) x 0.34

hemoglobin (g/dL) = hematocrit (decimal fraction) x 34

To convert a hemoglobin result from g/dL to mmol/L, multiply the displayed result by 0.621. The calculation of hemoglobin from hematocrit assumes a normal MCHC.

See below for information on factors affecting results. Certain substances, such as drugs, may affect analyte levels in vivo.² If results appear inconsistent with the clinical assessment, the patient sample should be retested using another cartridge.

REAGENTS

Contents

Each i-STAT cartridge contains one reference electrode sensor, sensors for the measurement of specific analytes, and a buffered aqueous calibrant solution that contains known concentrations of analytes and preservatives. A list of reactive ingredients for the EC4+ cartridge is shown below:

Sensor	Reactive Ingredient	Biological Source	Minimum Quantity
Na	Sodium (Na ⁺)	N/A	121 mmol/L
K	Potassium (K ⁺)	N/A	3.6 mmol/L
Glu	Glucose	N/A	7 mmol/L
	Glucose Oxidase	<i>Aspergillus niger</i>	0.002 IU

Warnings and Precautions

- For *in vitro* diagnostic use.
- Cartridges are intended for single-use only. Do not reuse.
- Refer to the i-STAT 1 System Manual for all warnings and precautions.

Storage Conditions

- Refrigeration at 2–8 °C (35–46 °F) until expiration date.
- Room Temperature at 18–30 °C (64–86 °F). Refer to the cartridge box for recommended shelf life.

INSTRUMENTS

The EC4+ cartridge is intended for use with i-STAT 1 analyzer.

SPECIMEN COLLECTION AND PREPARATION FOR ANALYSIS

Specimen Types

Arterial, venous or capillary whole blood.

Sample volume: 65 µL

Blood Collection Options and Test Timing (time from collection to cartridge fill)

Analyte	Syringes*	Test Timing	Evacuated Tubes	Test Timing	Capillary Tubes	Test Timing
Sodium Potassium Glucose Hematocrit	Without anticoagulant	3 minutes	Without anticoagulant	3 minutes	With balanced heparin anticoagulant or lithium heparin or lithium heparin if labeled for the measurement of electrolytes	3 minutes
	With balanced heparin anticoagulant or lithium heparin anticoagulant (syringe must be filled per manufacturer's recommendation) • Remix thoroughly before filling cartridge.	30 minutes	With lithium heparin anticoagulant (tubes must be filled per manufacturer's recommendation) • Remix thoroughly before filling cartridge.	30 minutes		

PROCEDURE FOR CARTRIDGE TESTING

Each cartridge is sealed in a foil pouch for protection during storage--do not use if pouch has been punctured.

- A cartridge should not be removed from its protective pouch until it is at room temperature (18-30 °C or 64-86 °F). For best results, the cartridge and analyzer should be at room temperature.
- Since condensation on a cold cartridge may prevent proper contact with the analyzer, allow refrigerated cartridges to equilibrate at room temperature for 5 minutes for a single cartridge and 1 hour for an entire box before use.
- Use a cartridge immediately after removing it from its protective pouch. Prolonged exposure may cause a cartridge to fail a Quality Check.
- Do not return unopened, previously refrigerated cartridges to the refrigerator.
- Cartridges may be stored at room temperature for the time frame indicated on the cartridge box.

Filling and Sealing the Cartridge (after cartridge has been equilibrated and blood sample has been collected)

1. Place the cartridge on a flat surface.
2. Mix the sample thoroughly. Invert a lithium heparin blood collection tube at least 10 times. If sample was collected into a syringe, invert syringe for 5 seconds then roll the syringe between the palms (hands parallel to the ground) for 5 seconds, flip and roll for an additional 5 seconds. The blood in the hub of the syringe will not mix, therefore expelling 2 drops before filling a cartridge is desired. Note that it may be difficult to properly mix a sample in a 1.0 mL syringe.
3. Fill the cartridge immediately after mixing. Direct the hub of syringe or tip of the transfer device (capillary tube, pipette, or dispensing tip) into the sample well of the cartridge.
4. Slowly dispense sample into the sample well until the sample reaches the fill mark indicated on the cartridge. Cartridge is properly filled when the sample reaches the 'fill to' mark and a small amount of sample is in the sample well. The sample should be continuous, no bubbles or breaks (see System Manual for details).
5. Fold the snap closure over the sample well.

Performing Patient Analysis

1. Press the power button to turn on the handheld.
2. Press 2 for *i-STAT Cartridge*.

3. Follow the handheld prompts.
4. Scan the lot number on the cartridge pouch.
5. Continue normal procedures for preparing the sample, and filling and sealing the cartridge.
6. Push the sealed cartridge into the handheld port until it clicks into place. Wait for the test to complete.
7. Review the results.

For additional information for cartridge testing, refer to the i-STAT 1 System Manual located at www.pointofcare.abbott.

Analysis Time

Approximately 130–200 seconds.

Quality Control

The i-STAT quality control regimen comprises four aspects, with a system design that reduces the opportunity for error, including:

1. A series of automated, on-line quality measurements that monitors the sensors, fluidics, and instrumentation each time a test is performed.
2. A series of automated, on-line procedural checks that monitors the user each time a test is performed.
3. Liquid materials are available to be used to verify the performance of a batch of cartridges when they are first received or when storage conditions are in question. The performance of this procedure is not a manufacturer's system instruction.
4. Traditional quality control measurements that verify the instrumentation using an independent device, which simulates the characteristics of the electrochemical sensors in a way that stresses the performance characteristics of the instrumentation.

For additional information on Quality Control, refer to the i-STAT 1 System Manual located at www.pointofcare.abbott.

Calibration Verification

Calibration Verification is a procedure intended to verify the accuracy of results over the entire measurement range of a test. The performance of this procedure is not a manufacturer's system instruction. However, it may be required by regulatory or accreditation bodies. While the Calibration Verification Set contains five levels, verification of the measurement range could be accomplished using the lowest, highest and mid-levels.

EXPECTED VALUES

TEST	UNITS *	REPORTABLE RANGE	REFERENCE RANGE	
			arterial	venous
MEASURED				
Na	mmol/L (mEq/L)	100–180	138–146 ³	
K	mmol/L (mEq/L)	2.0–9.0	3.5–4.9** ³	
Glu	mmol/L	1.1–38.9	3.9–5.8 ⁴	
	mg/dL	20–700	70–105 ⁴	
	g/L	0.20–7.00	0.70–1.05 ⁴	
Hematocrit/Hct	% PCV ***	15–75	38–51**** ³	
	Fraction	0.15–0.75	0.38–0.51 ³	
CALCULATED				
Hemoglobin/Hb	g/dL	5.1–25.5	12–17**** ³	
	g/L	51–255	120–170 ³	
	mmol/L	3.2–15.8	7–11 ³	

* The i-STAT System can be configured with the preferred units. Not applicable for pH test.

** The reference range for potassium has been reduced by 0.2 mmol/L from the range cited in Reference 3 to account for the difference in results between serum and plasma.

*** PCV, packed cell volume.

**** The reference ranges for hematocrit and hemoglobin span both female and male populations.

Unit Conversion

- **Glucose (Glu):** To convert mg/dL to mmol/L, multiply the mg/dL value by 0.055.
- **Hematocrit (Hct):** To convert a result from % PCV (packed cell volume) to fraction packed cell volume, divide the % PCV result by 100. For the measurement of hematocrit, the i-STAT System can be customized to agree with methods calibrated by the microhematocrit reference method using either K₃EDTA or K₂EDTA anticoagulant. Mean cell volumes of K₃EDTA anticoagulated blood are approximately 2–4% less than K₂EDTA anticoagulated blood. While the choice of anticoagulant affects the microhematocrit method to which all hematocrit methods are calibrated, results from routine samples on hematology analyzers are independent of the anticoagulant used. Since most clinical hematology analyzers are calibrated by the microhematocrit method using K₃EDTA anticoagulant, the i-STAT System default customization is K₃EDTA.

The reference ranges programmed into the analyzer and shown above are intended to be used as guides for the interpretation of results. Since reference ranges may vary with demographic factors such as age, gender and heritage, it is recommended that reference ranges be determined for the population being tested.

METROLOGICAL TRACEABILITY

The measured analytes in the i-STAT EC4+ cartridge are traceable to the following reference materials or methods. The i-STAT System controls and calibration verification materials are validated for use only with the i-STAT System and assigned values may not be commutable with other methods.

Sodium (Na) and Potassium (K)

The respective analyte values assigned to i-STAT System controls and calibration verification materials are traceable to the U.S. National Institute of Standards and Technology (NIST) standard reference

material SRM956.

Glucose (Glu)

The i-STAT System test for glucose measures glucose amount-of-substance concentration in the plasma fraction of arterial, venous, or capillary whole blood (dimension mmol L⁻¹) for *in vitro* diagnostic use. Glucose values assigned to i-STAT System controls and calibration verification materials are traceable to the U.S. National Institute of Standards and Technology (NIST) standard reference material SRM965.

Hematocrit (Hct)

The i-STAT System test for hematocrit measures packed red blood cell volume fraction in arterial, venous, or capillary whole blood (expressed as the % packed cell volume) for *in vitro* diagnostic use. Hematocrit values assigned to i-STAT System working calibrators are traceable to the Clinical and Laboratory Standards Institute (CLSI) H7-A3 procedure for determining packed cell volume by the microhematocrit method.⁵

Additional information regarding metrological traceability is available from Abbott Point of Care Inc.

PERFORMANCE CHARACTERISTICS

The typical performance data summarized below were collected in health care facilities by health care professionals trained in the use of the i-STAT System and comparative methods.

Precision

Precision data collected was collected in multiple sites and tested as follows: Duplicates of each control fluid were tested in the morning and in the afternoon on five days for a total of 20 replicates. The averaged statistics are presented below.

Test	Units	Aqueous Control	Mean	SD (Standard Deviation)	CV (%) [Coefficient of Variation (%)]
Na	mmol/L or mEq/L	Level 1	120.0	0.46	0.4
		Level 3	160.0	0.53	0.3
K	mmol/L or mEq/L	Level 1	2.85	0.038	1.3
		Level 3	6.30	0.039	0.6
Glu	mg/dL	Level 1	41.8	0.68	1.6
		Level 3	289	2.4	0.8
Hct	% PCV (packed cell volume)	Low	30.0	0.44	1.5
		High	49.0	0.50	1.0

Method Comparison

Method comparison data was collected using CLSI guideline EP9-A.⁶

Deming regression analysis⁷ was performed on the first replicate of each sample set. In the method comparison table, n is the number of specimens in the data set, Sxx and Syy refer to estimates of imprecision based on the duplicates of the comparative and the i-STAT methods respectively, Sy.x is the standard error of the estimate, and r is the correlation coefficient.*

Method comparisons will vary from site to site due to differences in sample handling, comparative method calibration and other site-specific variables.

* The usual warning relating to the use of regression analysis is summarized here as a reminder. For any analyte, "if the data is collected over a narrow range, the estimate of the regression parameters are relatively imprecise and may be biased. Therefore, predictions made from these estimates may be invalid".⁷ The correlation coefficient, r, can be

used as a guide to assess the adequacy of the comparative method range in overcoming this problem, and, as a guide, the range of data can be considered adequate for $r > 0.975$.

Sodium/Na (mmol/L or mEq/L)		Beckman Synchron CX®3	Kodak Ektachem™ 700	Nova STAT Profile® 5	
Venous blood samples were collected in lithium heparin Vacutainer® tubes and analyzed in duplicate on the i-STAT System. A portion of the specimen was centrifuged and the separated plasma was analyzed in duplicate on comparative methods within 20 minutes of collection.	n	189	142	192	
	Sxx	0.74	0.52	0.54	
	Syy	0.53	0.58	0.53	
	Slope	1.00	0.98	0.95	
	Int't	-0.11	3.57	5.26	
	Sy.x	1.17	1.04	1.53	
	Xmin	126	120	124	
	Xmax	148	148	148	
	r	0.865	0.937	0.838	
Potassium/K (mmol/L or mEq/L)		Beckman Synchron CX®3	Kodak Ektachem™ 700	Nova STAT Profile® 5	
Venous blood samples were collected in lithium heparin Vacutainer® tubes and analyzed in duplicate on the i-STAT System. A portion of the specimen was centrifuged and the separated plasma was analyzed in duplicate on comparative methods within 20 minutes of collection.	n	189	142	192	
	Sxx	0.060	0.031	0.065	
	Syy	0.055	0.059	0.055	
	Slope	0.97	1.06	0.99	
	Int't	0.02	-0.15	-0.01	
	Sy.x	0.076	0.060	0.112	
	Xmin	2.8	3.0	2.8	
	Xmax	5.7	9.2	5.8	
	r	0.978	0.993	0.948	
Glucose/Glu (mg/dL)		Beckman Coulter LX20®	Bayer 860	Dade Dimension RxL-Xpand	
Venous blood samples were collected in lithium heparin Vacutainer® tubes and analyzed in duplicate on the i-STAT System. A portion of the specimen was centrifuged and the separated plasma was analyzed in duplicate on comparative methods within 20 minutes of collection.	n	35	40	32	
	Sxx	2.21	4.71	0.98	
	Syy	0.69	0.96	0.59	
	Slope	1.03	0.99	1.01	
	Int't	-3.39	-1.67	-0.85	
	Sy.x	0.91	0.70	1.57	
	Xmin	45	58	48	
	Xmax	297	167	257	
	r	0.999	0.993	0.998	
Hematocrit/Hct (% PCV) (% packed cell volume)		Coulter® S Plus	Nova STAT Profile® 5	Abbott Cell-Dyn 4000	Sysmex SE9500
Venous blood samples, collected in lithium heparin Vacutainer® tubes, were analyzed in duplicate on the i-STAT System and on the comparative methods for hematocrit within 20 minutes of collection.	n	142	192	29	29
	Sxx	0.50	0.46	0.41	0.53
	Syy	1.09	1.31	0.77	0.76
	Slope	0.98	1.06	1.06	1.11
	Int't	1.78	-3.98	-1.42	-4.19
	Sy.x	2.03	2.063	1.13	0.98
	Xmin	18	21	19	24
	Xmax	51	50	46	47

	r	0.952	0.932	0.993	0.980
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FACTORS AFFECTING RESULTS

The following substances were evaluated in plasma for relevant analytes at the test concentrations recommended in CLSI guideline EP7-A2 ⁸ unless otherwise noted. For those identified as an interferant the interference is described.

Substance	Test Concentration (mmol/L)	Analyte	Interference (Yes/No)	Comment
Acetaldehyde	0.045 ⁹	Glu	No	
Acetaminophen	1.32	Na	No	
		K	No	
		Glu	Yes	Increased results
Acetaminophen (therapeutic)	0.132 ⁹	Glu	No	
Acetoacetate	2.0	Glu	No	
Acetylcysteine	10.2	Na	No	
		K	No	
		Glu	Yes	Decreased results
Acetylcysteine (therapeutic)	0.30 ^{10 11}	Glu	No	
Ascorbate	0.34	Na	No	
		K	No	
		Glu	No	
Bromide	37.5	Na	Yes	Increased results. Use another method.
		K	Yes	Increased results and rate of star (***) outs. Use another method.
		Glu	Yes	Decreased results. Use another method.
		Hct	Yes	Increased rate of star (***) outs
Bromide (therapeutic)	2.5 ^{12 13 14}	Na	No	
		K	No	
		Glu	Yes	Decreased results
		Hct	No	
Dopamine	0.006	Glu	No	
Formaldehyde	0.133 ⁹	Glu	No	
β-Hydroxybutyrate	6.0 ¹⁵	Na	No	
		K	No	
		Glu	No	

Substance	Test Concentration (mmol/L)	Analyte	Interference (Yes/No)	Comment
Hydroxyurea	0.92	Glu	Yes	Increased results. Use another method.
Lactate	6.6	Na	No	
		K	No	
		Glu	No	
Magnesium Chloride	1.0	Na	No	
		K	No	
Maltose	13.3	Glu	No	
Nithiodote (Sodium thiosulfate)	16.7 ¹⁶	Na	Yes	Increased results
		K	Yes	Decreased results
		Glu	Yes	Decreased results
Pyruvate	0.31	Glu	No	
Salicylate	4.34	Na	No	
		K	No	
		Glu	No	
Thiocyanate	6.9	Glu	Yes	Decreased results
Thiocyanate (therapeutic)	0.5 ⁹	Glu	No	
Uric Acid	1.4	Glu	No	

The degree of interference at concentrations other than those reported above might not be predictable. It is possible that interfering substances other than those tested may be encountered.

Relevant comments regarding interference of Acetaminophen, Acetylcysteine, Bromide, Hydroxyurea and Nithiodote are noted below:

- Acetaminophen has been shown to interfere with i-STAT glucose results at a concentration proscribed by the CLSI guideline, 1.32 mmol/L, which represents a toxic concentration. Acetaminophen at 0.132 mmol/L, which represents the upper end of the therapeutic concentration, has been shown not to significantly interfere with i-STAT glucose results.
- Acetylcysteine has been tested at two levels: the CLSI recommended level of 10.2 mmol/L and a concentration of 0.30 mmol/L. The latter is 3 times the peak therapeutic plasma concentration associated with treatment to reverse acetaminophen poisoning. APOC has not identified a therapeutic condition that would lead to levels consistent with the CLSI recommended level.
- Bromide has been tested at two levels: the CLSI recommended level and a therapeutic plasma concentration level of 2.5 mmol/L. The latter is the peak plasma concentration associated with halothane anesthesia, in which bromide is released. APOC has not identified a therapeutic condition that would lead to levels consistent with the CLSI recommended level.
- Hydroxyurea has been shown to interfere with glucose results at 0.92 mmol/L. Hydroxyurea is a DNA synthesis inhibitor used in the treatment sickle cell anemia, HIV infection, and various types of cancer. The malignancies that it is used to treat include melanoma, metastatic ovarian cancer, and chronic myelogenous leukemia. It is also used in the treatment of polycythemia vera, thrombocythemia, and psoriasis. At typical doses ranging from 500 mg to 2 g/day, concentrations of hydroxyurea in a patient's blood may be sustained at approximately 100 to 500 µmol/L. Higher concentrations may be observed soon after dosing or at higher therapeutic doses.
- Nithiodote (sodium thiosulfate) has been shown to interfere with sodium, potassium and glucose














results at 16.7 mmol/L. Nithiodote (sodium thiosulfate) is indicated for the treatment of acute cyanide poisoning. The journal article titled “Falsely increased chloride and missed anion gap elevation during treatment with sodium thiosulfate” indicated that sodium thiosulfate could be used in the treatment of calciphylaxis indicating that “the highest concentration likely to be seen in plasma [is] after infusion of a 12.5 g dose of sodium thiosulfate pentahydrate. Assuming that the 12.5 g dose of sodium thiosulfate pentahydrate is distributed in a typical blood volume of 5 L with a hematocrit of 40%, the peak sodium thiosulfate plasma concentration expected is 16.7 mmol/L.”¹⁶

OTHER FACTORS AFFECTING RESULTS

Factor	Analyte	Effect
Sodium heparin	Na	Sodium heparin may increase sodium results up to 1 mmol/L. ¹⁷
Hemodilution	Na	Hemodilution of the plasma by more than 20% associated with priming cardiopulmonary bypass pumps, plasma volume expansion or other fluid administration therapies using certain solutions may cause clinically significant error on sodium and chloride results. These errors are associated with solutions that do not match the ionic characteristics of plasma. To minimize these errors when hemodiluting by more than 20%, use physiologically balanced multi-electrolyte solutions containing low-mobility anions (e.g., gluconate).
Line draw	Hct	Low hematocrit results can be caused by contamination of flush solutions in arterial or venous lines. Back flush a line with a sufficient amount of blood to remove intravenous solutions, heparin, or medications that may contaminate the sample. Five to six times the volume of the catheter, connectors, and needle is recommended.
Cold temperature	K	Potassium values will increase in iced specimens.
Allowing blood to stand (without exposure to air)	K	If heparinized whole blood is allowed to stand before testing, potassium values will first decrease slightly, then increase over time.
	Glu	Glucose values will decrease in whole blood samples over time. Venous blood glucose is as much as 7 mg/dL less than capillary blood glucose as a result of tissue utilization. ¹⁸
Sample type	K	Serum Potassium results may be 0.1 to 0.7 mmol/L higher than Potassium results from anticoagulated samples due to the release of Potassium from platelets ¹ and red blood cells during the clotting process.
Sample mixing	Hct	Samples from 1 mL syringes should not be used to determine hematocrit if testing is delayed.
Hemolysis	K	Potassium values obtained from skin puncture samples may vary due to hemolysis or an increase in tissue fluid from improper technique during the collection procedure.
pH dependence	Glu	The dependence of the i-STAT glucose test with respect to pH is as follows: values below pH 7.4 at 37°C decrease results by approximately 0.9 mg/dL (0.05 mmol/L) per 0.1 pH unit. Values above pH 7.4 at 37 °C increase results by approximately 0.8 mg/dL (0.04 mmol/L) per 0.1 pH unit.
PO ₂ dependence	Glu	The dependence of the i-STAT glucose test with respect to PO₂ is as follows: oxygen levels of less than 20 mmHg (2.66 kPa) at 37°C may decrease results.
Erythrocyte	Hct	<ul style="list-style-type: none"> The measurement of certain blood samples with high erythrocyte

Factor	Analyte	Effect									
sedimentation rate		<p>sedimentation rates (ESR) may be affected by analyzer angle. While testing blood samples, beginning 90 seconds after the cartridge is inserted, the analyzer should remain level until a result is obtained. A level surface includes running the handheld in the downloader/ recharger.</p> <ul style="list-style-type: none"> Hematocrit results can be affected by the settling of red blood cells in the collection device. The best way to avoid the effect of settling is to test the sample immediately. If there is a delay in testing of a minute or more, the sample must be remixed thoroughly. 									
White Blood Cell Count (WBC)	Hct	Grossly elevated white blood cell counts may increase results.									
Lipids	Hct	Abnormally high lipids may increase results. Interference from lipids will be about two thirds the size of the interference from protein.									
Total Protein	Hct	<p>Hematocrit results are affected by the level of total protein as follows:</p> <table border="1"> <thead> <tr> <th>Displayed Result</th><th>Total Protein (TP) < 6.5 g/dL</th><th>Total Protein (TP) > 8.0 g/dL</th></tr> </thead> <tbody> <tr> <td>HCT < 40% PCV</td><td>Hct decreased by ~1% PCV for each decrease of 1 g/dL TP</td><td>Hct increased by ~1% PCV for each increase of 1 g/dL TP</td></tr> <tr> <td>HCT > 40% PCV</td><td>Hct decreased by ~0.75 % PCV for each decrease of 1 g/dL TP</td><td>Hct increased by ~0.75 % PCV for each increase of 1 g/dL TP</td></tr> </tbody> </table> <ul style="list-style-type: none"> Total protein levels may be low in neonatal and burn patient populations, as well as in additional clinical populations listed in Statland.3 Total protein levels may also be decreased in patients undergoing cardiopulmonary bypass (CPB) or extracorporeal membrane oxygenation (ECMO) and with patients receiving large volumes of saline-based intravenous (IV) fluids. Care should be taken when using hematocrit results from patients with total protein levels below the adult reference range (6.5 to 8 g/dL). The CPB sample type can be used to correct the hematocrit result for the dilutional effect of the pump prime in cardiovascular surgery. The CPB algorithm assumes that cells and plasma are diluted equally and that the pump priming solution has no added albumin or other colloid or packed red blood cells. Since perfusion practices vary, it is recommended that each practice verify the use of the CPB sample type and the length of time in which the CPB sample type should be used during the recovery period. Note that for hematocrit values above 30% PCV, the CPB correction is $\leq 1.5\%$ PCV; the size of the correction at this level should not impact transfusion decisions. 	Displayed Result	Total Protein (TP) < 6.5 g/dL	Total Protein (TP) > 8.0 g/dL	HCT < 40% PCV	Hct decreased by ~1% PCV for each decrease of 1 g/dL TP	Hct increased by ~1% PCV for each increase of 1 g/dL TP	HCT > 40% PCV	Hct decreased by ~0.75 % PCV for each decrease of 1 g/dL TP	Hct increased by ~0.75 % PCV for each increase of 1 g/dL TP
Displayed Result	Total Protein (TP) < 6.5 g/dL	Total Protein (TP) > 8.0 g/dL									
HCT < 40% PCV	Hct decreased by ~1% PCV for each decrease of 1 g/dL TP	Hct increased by ~1% PCV for each increase of 1 g/dL TP									
HCT > 40% PCV	Hct decreased by ~0.75 % PCV for each decrease of 1 g/dL TP	Hct increased by ~0.75 % PCV for each increase of 1 g/dL TP									
Sodium	Hct	The sample electrolyte concentration is used to correct the measured conductivity prior to reporting hematocrit results. Factors that affect sodium will therefore also affect hematocrit.									

KEY TO SYMBOLS

Symbol	Definition/Use
	14 days room temperature storage at 18–30 °C.
	Use by or expiration date. The expiration date, expressed as YYYY-MM-DD, indicates the last day the product may be used.
	Manufacturer's lot number or batch code. The lot number or batch code appears adjacent to this symbol.
	Sufficient for <n> tests.
	Authorized representative for Regulatory Affairs in the European Community.
	Temperature limitations. The upper and lower limits for storage are adjacent to upper and lower arms.
	Catalog number, list number, or reference.
	Do not reuse.
	Manufacturer.
	Consult instructions for use or see System Manual for instructions.
	<i>In vitro</i> diagnostic medical device.
	Compliance to the European directive on <i>in vitro</i> diagnostic devices (98/79/EC)
	For prescription use only.

Additional Information: to obtain additional product information and technical support, refer to the Abbott company website at www.pointofcare.abbott.

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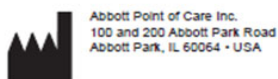
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