

# i-STAT G3+ Cartridge

Intended for use with the i-STAT Alinity Instrument

## NAME

i-STAT G3+ Cartridge – REF 03P78-25



## INTENDED USE

The i-STAT G3+ cartridge with the i-STAT Alinity System is intended for use in the *in vitro* quantification of pH, partial pressure of oxygen, and partial pressure of carbon dioxide in arterial, venous or capillary whole blood.

Analyte	Intended Use
pH	pH, $PO_2$ , and $PCO_2$ measurements are used in the diagnosis, monitoring, and treatment of respiratory disturbances and metabolic and respiratory-based acid-base disturbances.
Partial Pressure of Oxygen ( $PO_2$ )	
Partial Pressure of Carbon Dioxide ( $PCO_2$ )	Bicarbonate is used in the diagnosis and treatment of numerous potentially serious disorders associated with changes in body acid-base balance.

## SUMMARY AND EXPLANATION/CLINICAL SIGNIFICANCE

### Measured:

#### pH

pH is an index of the acidity or alkalinity of the blood with an arterial pH of  $<7.35$  indicating an acidemia and  $>7.45$  alkalemia. <sup>1</sup>

#### Partial Pressure of Oxygen ( $PO_2$ )

$PO_2$  (partial pressure of oxygen) is a measurement of the tension or pressure of oxygen dissolved in blood. Some causes for decreased values of  $PO_2$  include decreased pulmonary ventilation (e.g., airway obstruction or trauma to the brain), impaired gas exchange between alveolar air and pulmonary capillary blood (e.g., bronchitis, emphysema, or pulmonary edema), and alteration in the flow of blood within the heart or lungs (e.g., congenital defects in the heart or shunting of venous blood into the arterial system without oxygenation in the lungs).

#### Partial Pressure of Carbon Dioxide ( $PCO_2$ )

$PCO_2$  along with pH is used to assess acid-base balance.  $PCO_2$  (partial pressure of carbon dioxide), the respiratory component of acid-base balance, is a measure of the tension or pressure of carbon dioxide dissolved in the blood.  $PCO_2$  represents the balance between cellular production of  $CO_2$  and ventilatory removal of  $CO_2$  and a change in  $PCO_2$  indicates an alteration in this balance. Causes of primary respiratory acidosis (increase in  $PCO_2$ ) are airway obstruction, sedatives and anesthetics, respiratory distress syndrome, and chronic obstructive pulmonary disease. Causes of primary respiratory alkalosis (decreased  $PCO_2$ ) are hypoxia (resulting in hyperventilation) due to chronic heart failure, edema and neurologic disorders, and mechanical hyperventilation.

## TEST PRINCIPLE

The i-STAT System uses direct (undiluted) electrochemical methods. Values obtained by direct methods may differ from those obtained by indirect (diluted) methods. <sup>2</sup>

## Measured:

### pH

pH is measured by direct potentiometry. In the calculation of results for pH, concentration is related to potential through the Nernst equation.

### PO<sub>2</sub>

PO<sub>2</sub> is measured amperometrically. The oxygen sensor is similar to a conventional Clark electrode. Oxygen permeates through a gas permeable membrane from the blood sample into an internal electrolyte solution where it is reduced at the cathode. The oxygen reduction current is proportional to the dissolved oxygen concentration.

### PCO<sub>2</sub>

PCO<sub>2</sub> is measured by direct potentiometry. In the calculation of results for PCO<sub>2</sub>, concentration is related to potential through the Nernst equation.

### Temperature “Correction” Algorithm

pH, PO<sub>2</sub>, and PCO<sub>2</sub> are temperature-dependent quantities and are measured at 37°C. The pH, PO<sub>2</sub>, and PCO<sub>2</sub> readings at a body temperature other than 37°C can be ‘corrected’ by entering the patient’s temperature on the chart page of the analyzer. In this case, blood gas results will be displayed at both 37°C and the patient’s temperature.

pH, PO<sub>2</sub>, and PCO<sub>2</sub> at the patient’s temperature (T<sub>p</sub>) are calculated as follows:<sup>3</sup>

$$pH(T_p) = pH - 0.0147(T_p - 37) + 0.0065(7.4 - pH)(T_p - 37)$$

$$PO_2(T_p) = PO_2 \times 10^{\frac{5.49 \times 10^{-11} PO_2^{3.88} + 0.071}{9.72 \times 10^{-9} PO_2^{3.88} + 2.30} (T_p - 37)}$$

$$PCO_2(T_p) = PCO_2 \times 10^{0.019(T_p - 37)}$$

## Calculated:

### HCO<sub>3</sub>, TCO<sub>2</sub>, and BE

- HCO<sub>3</sub> (bicarbonate), the most abundant buffer in the blood plasma, is an indicator of the buffering capacity of blood. Regulated primarily by the kidneys, HCO<sub>3</sub> is the metabolic component of acid-base balance.
- TCO<sub>2</sub> is a measure of carbon dioxide which exists in several states: CO<sub>2</sub> in physical solution or loosely bound to proteins, bicarbonate (HCO<sub>3</sub>) or carbonate (CO<sub>3</sub>) anions, and carbonic acid (H<sub>2</sub>CO<sub>3</sub>). Measurement of TCO<sub>2</sub> as part of an electrolyte profile is useful chiefly to evaluate HCO<sub>3</sub> concentration. TCO<sub>2</sub> and HCO<sub>3</sub> are useful in the assessment of acid-base imbalance (along with pH and PCO<sub>2</sub>) and electrolyte imbalance.
- The calculated TCO<sub>2</sub> provided by the i-STAT System is determined from the measured and reported values of pH and PCO<sub>2</sub> according to a simplified and standardized form of the Henderson-Hasselbalch equation.<sup>3</sup>
- This calculated TCO<sub>2</sub> measurement is metrologically traceable to the i-STAT pH and PCO<sub>2</sub> measurements, which are in turn traceable to primary standard reference materials for pH and PCO<sub>2</sub>. Like all calculated parameters reported by the i-STAT System, the user can independently determine TCO<sub>2</sub> values from the reported pH and PCO<sub>2</sub> measurements using a combination of the equation for HCO<sub>3</sub> given in the PCO<sub>2</sub>.

- Base excess of the extracellular fluid (ECF) or standard base excess is defined as the concentration of titratable base minus the concentration of titratable acid when titrating the average ECF (plasma plus interstitial fluid) to an arterial plasma pH of 7.40 at  $PCO_2$  of 40 mmHg at 37 °C. Excess concentration of base in the average ECF remains virtually constant during acute changes in the  $PCO_2$  and reflects only the non-respiratory component of pH-disturbances.

When a cartridge includes sensors for both pH and  $PCO_2$ , bicarbonate ( $HCO_3$ ), total carbon dioxide ( $TCO_2$ ) and base excess (BE) are calculated.<sup>3</sup>

$$\begin{aligned} \log HCO_3 &= pH + \log PCO_2 - 7.608 \\ TCO_2 &= HCO_3 + 0.03 PCO_2 \\ BE_{ecf} &= HCO_3 - 24.8 + 16.2(pH - 7.4) \\ BE_b &= (1 - 0.014 * Hb) * [ HCO_3 - 24.8 + (1.43 * Hb + 7.7) * (pH - 7.4) ] \end{aligned}$$

## sO<sub>2</sub>

- sO<sub>2</sub> (oxygen saturation) is the amount of oxyhemoglobin expressed as a fraction of the total amount of hemoglobin able to bind oxygen (oxyhemoglobin plus deoxyhemoglobin).
- sO<sub>2</sub> is calculated from measured  $PO_2$  and pH and from  $HCO_3$  calculated from measured  $PCO_2$  and pH. However, this calculation assumes normal affinity of oxygen for hemoglobin. It does not take into account erythrocyte diphosphoglycerate (2,3-DPG) concentrations which affect the oxygen dissociation curve. The calculation also does not take into account the effects of fetal hemoglobin or dysfunctional hemoglobins (carboxy-, met-, and sulfhemoglobin). Clinically significant errors can result from incorporation of such an estimated sO<sub>2</sub> value for oxygen saturation in further calculations, such as shunt fraction, or by assuming the value obtained is equivalent to fractional oxyhemoglobin.

$$sO_2 = 100 \frac{(X^3 + 150X)}{X^3 + 150X + 23400}$$

where  $X = PO_2 \cdot 10^{(0.48(pH-7.4) - 0.0013(HCO_3 - 25))}$

See below for information on factors affecting results. Certain substances, such as drugs, may affect analyte levels in vivo.<sup>4</sup> If results appear inconsistent with the clinical assessment, the patient sample should be retested using another cartridge.

## REAGENTS

### Contents

Each i-STAT cartridge contains one reference electrode, sensors for the measurement of specific analytes, and a buffered aqueous calibrant solution that contains known concentrations of analytes and preservatives. A list of reactive ingredients relevant for the i-STAT G3+ cartridge is indicated below:

Sensor	Reactive Ingredient	Biological Source	Minimum Quantity
pH	Hydrogen Ion (H <sup>+</sup> )	N/A	6.66 pH
$PCO_2$	Carbon Dioxide (CO <sub>2</sub> )	N/A	25.2 mmHg

### Warnings and Precautions

- For *in vitro* diagnostic use.
- Cartridges are intended for single-use only. Do not reuse.
- Refer to the i-STAT Alinity System Operations Manual for all warnings and precautions.

### Storage Conditions

- Refrigeration at 2–8 °C (35–46 °F) until expiration date.
- Room Temperature at 18–30 °C (64–86 °F). Refer to the cartridge box for room temperature storage requirements.

## INSTRUMENTS

The i-STAT G3+ cartridge is intended for use with the i-STAT Alinity Instrument (Model No. AN-500).

## SPECIMEN COLLECTION AND PREPARATION FOR ANALYSIS

### Specimen Types

Arterial, venous, capillary whole blood.

Sample Volume: 95 µL

### Blood Collection Options and Test Timing (time from collection to cartridge fill)

**As higher heparin-to-blood ratios may affect results, fill blood collections tubes and syringes to capacity, following manufacturers' instructions.**

<b>G3+ Sample Collection</b>	
Syringe	<b>Without anticoagulant</b> <ul style="list-style-type: none"><li>• Maintain anaerobic conditions prior to filling this cartridge.</li><li>• Mix sample immediately before filling cartridge.</li><li>• Fill cartridge within 3 minutes of sample collection.</li></ul> <b>With balanced heparin anticoagulant</b> <ul style="list-style-type: none"><li>• Maintain anaerobic conditions prior to filling this cartridge.</li><li>• Mix sample immediately before filling cartridge.</li><li>• Fill cartridge within 10 minutes of sample collection.</li></ul>
Evacuated Tube	<b>Without anticoagulant</b> <ul style="list-style-type: none"><li>• Maintain anaerobic conditions prior to filling this cartridge.</li><li>• Mix sample immediately before filling cartridge.</li><li>• Fill cartridge within 3 minutes of sample collection.</li></ul> <b>With lithium heparin anticoagulant</b> <ul style="list-style-type: none"><li>• Maintain anaerobic conditions prior to filling this cartridge.</li><li>• Mix sample immediately before filling cartridge.</li><li>• Fill cartridge within 10 minutes of sample collection</li></ul>
Capillary Tube	<b>With balanced heparin anticoagulant</b> <ul style="list-style-type: none"><li>• Mix sample immediately before filling cartridge.</li><li>• Fill cartridge within 3 minutes of sample collection.</li></ul> <b>With lithium heparin anticoagulant</b> <ul style="list-style-type: none"><li>-If labeled for measurement of electrolytes.</li><li>• Mix sample immediately before filling cartridge.</li><li>• Fill cartridge within 3 minutes of sample collection</li></ul>
Fill cartridge directly from skin puncture	<b>Not Recommended</b>

## PROCEDURE FOR CARTRIDGE TESTING

Preparation for Use:

1. Individual cartridges may be used after standing five minutes at room temperature. An entire box of cartridges should stand at room temperature for one hour.
2. All cartridges should be used immediately after opening pouch.
3. If the pouch has been punctured, the cartridge should not be used.
4. Do not return cartridges to the refrigerator after bringing them to room temperature.

### How to Perform Patient Testing

1. From the Home screen, touch "**Perform Patient Test**". This initiates the patient testing pathway.
2. To begin, follow instructions on the screen to "**Scan or Enter OPERATOR ID**"
3. Follow instructions on the screen to "**Scan or Enter PATIENT ID**"

4. Continue to follow prompts on the screen to proceed with patient testing. **“Scan (CARTRIDGE POUCH) Barcode”**, Scanning is required. Information cannot be entered manually.
5. The screen for selecting sample type will display if more than one sample type is applicable; select sample type if applicable.
6. Follow instructions on the screen to **“Close and Insert Filled Cartridge”**. The action buttons at the bottom of the screen allow forward, backward and pause functionality.
7. Once the cartridge is inserted, **“Contacting Cartridge”** will display followed by the countdown bar. The following alerts are also displayed: **“Cartridge locked in instrument. Do not attempt to remove the Cartridge”** and **“Testing - Instrument Must Remain Level”**.
8. When the test is complete, the test results are displayed.

### **Analysis Time**

Approximately 130–200 seconds.

### **Quality Control**

The i-STAT Alinity System quality control regimen comprises various aspects, with a system design that reduces the opportunity for error, including:

1. The i-STAT Alinity System automatically runs a comprehensive set of quality checks of analyzer and cartridge performance each time a sample is tested. This internal quality system will suppress results if the analyzer or cartridge does not meet certain internal specifications.
2. Aqueous-based control solutions are available for verifying the integrity of newly received cartridges.
3. In addition, the instrument performs internal electronic checks and calibration during each test cycle, and the Electronic Simulator test provides an independent check on the ability of the instrument to take accurate and sensitive measurements of voltage, current and resistance from the cartridge. The instrument will pass or fail this electronic test depending on whether or not it measures these signals within limits specified in the instrument software.

For additional information on Quality Control, refer to the i-STAT Alinity System Operations Manual located at [www.pointofcare.abbott](http://www.pointofcare.abbott).

### **Calibration Verification**

Standardization is the process by which a manufacturer establishes “true” values for representative samples. A multi-point calibration is derived for each sensor by this standardization process. These calibration curves are stable over many lots.

A one-point calibration is performed each time a cartridge requiring calibration is used. During the first part of the testing cycle, the calibrant solution is automatically released from its foil pack and is positioned over the sensors. The signals produced by the sensors’ responses to the calibrant solution are measured. This one-point calibration adjusts the offset of the stored calibration curve. Next, the instrument automatically moves the sample over the sensors and the signals produced by the sensors’ responses to the sample are measured. While coefficients are used rather than graphic calibration curves, the calculation of the result is equivalent to reading the sample’s concentration from an adjusted calibration curve.

## EXPECTED VALUES

TEST	UNITS *	REPORTABLE RANGE	REFERENCE RANGE	
			(arterial)	(venous)
<b>MEASURED</b>				
pH		6.50 - 8.20	7.35 - 7.45 <sup>5</sup>	7.31 - 7.41 <sup>**</sup>
<i>PO</i> <sub>2</sub>	mmHg	5 - 800	80 - 105 <sup>6***</sup>	
	kPa	0.7 - 106.6	10.7 - 14.0 <sup>6***</sup>	
<i>PCO</i> <sub>2</sub>	mmHg	5 - 130	35 - 45 <sup>5</sup>	41 - 51
	kPa	0.67 - 17.33	4.67 - 6.00	5.47 - 6.80
<b>CALCULATED</b>				
Bicarbonate/ HCO <sub>3</sub>	mmol/L (mEq/L)	1.0 - 85.0	22 - 26 <sup>**</sup>	23 - 28 <sup>**</sup>
TCO <sub>2</sub>	mmol/L (mEq/L)	5 - 50	23 - 27	24 - 29
Base Excess/ BE	mmol/L (mEq/L)	(-30) - (+30)	(-2) - (+3) <sup>5</sup>	(-2) - (+3) <sup>5</sup>
sO <sub>2</sub>	%	0-100	95 - 98	

\* The i-STAT System can be configured with the preferred units. Not applicable for pH test.

\*\* Calculated from Siggard-Andersen nomogram.<sup>1</sup>

\*\*\* The reference ranges shown are for a healthy population. Interpretation of blood gas measurements depend on the underlying condition (e.g., patient temperature, ventilation, posture and circulatory status).

### Unit Conversion:

- *PO*<sub>2</sub> and *PCO*<sub>2</sub>: To convert *PO*<sub>2</sub> and *PCO*<sub>2</sub> results from mmHg to kPa, multiple the mmHg value by 0.133.

i-STAT Alinity does not have default reference ranges programmed into the instrument. The reference ranges shown above are intended to be used as guides for the interpretation of results. Since reference ranges may vary with demographic factors such as age, gender and heritage, it is recommended that reference ranges be determined for the population being tested.

## METROLOGICAL TRACEABILITY

The measured analytes in the i-STAT G3+ cartridge are traceable to the following reference materials or methods. The i-STAT System controls and calibration verification materials are validated for use only with the i-STAT System and assigned values may not be commutable with other methods.

### pH

The i-STAT System test for pH measures the hydrogen ion amount-of-substance concentration in the plasma fraction of arterial, venous or capillary whole blood (expressed as the negative logarithm of the relative molal hydrogen ion activity) for *in vitro* diagnostic use. pH values assigned to i-STAT System controls and calibration verification materials are traceable to the U.S. National Institute of Standards and Technology (NIST) standard reference materials SRMs 186-I, 186-II, 185, and 187.

### *PO*<sub>2</sub>

The i-STAT System test for oxygen partial pressure of oxygen measures the partial pressure of oxygen in arterial, venous or capillary whole blood (dimension kPa) for *in vitro* diagnostic use. *PO*<sub>2</sub> values assigned to i-STAT System controls and calibration verification materials are traceable to U.S. National Institute of Standards and Technology (NIST) standard reference materials via commercially available certified specialty medical gas standards.

## PCO<sub>2</sub>

The i-STAT System test for partial pressure of carbon dioxide measures the partial pressure of carbon dioxide in arterial, venous or capillary whole blood (dimension kPa) for *in vitro* diagnostic use. PCO<sub>2</sub> values assigned to i-STAT System controls and calibration verification materials are traceable to U.S. National Institute of Standards and Technology (NIST) standard reference materials via commercially available certified specialty medical gas standards.

Additional information regarding metrological traceability is available from Abbott Point of Care Inc.

## PERFORMANCE CHARACTERISTICS

The typical performance data summarized below was collected in health care facilities by health care professionals trained in the use of the i-STAT System and comparative methods.

### Precision\*

Precision data was collected in multiple sites as follows: Duplicates of each control fluid were tested in the morning and in the afternoon on five days for a total of 20 replicates. The averaged statistics are presented below.

Test	Units	Aqueous Cal Ver	n	Mean	SD (Standard Deviation)	CV (%) [Coefficient of Variation (%)]
pH		Very Low Abnormal	80	6.562	0.005	0.08
		Low Abnormal	80	7.031	0.004	0.06
		Normal	80	7.469	0.003	0.04
		High Abnormal	80	7.769	0.003	0.04
		Very High Abnormal	80	7.986	0.004	0.05
PO <sub>2</sub>	mmHg	Very Low Abnormal	80	72.1	2.02	2.80
		Low Abnormal	80	84.2	1.60	1.90
		Normal	80	118.8	2.10	1.77
		High Abnormal	80	152.1	3.49	2.29
		Very High Abnormal	80	377.1	8.52	2.26
PCO <sub>2</sub>	mmHg	Very Low Abnormal	80	17.4	0.43	2.5
		Low Abnormal	80	21.7	0.40	1.8
		Normal	80	28.7	0.57	2.0
		High Abnormal	80	56.2	1.18	2.1
		Very High Abnormal	80	84.5	1.93	2.3

\*Note: Representative data, individual laboratories may vary from these data.

### Method Comparison

Method comparison was demonstrated in a study comparing the i-STAT Alinity to the i-STAT 1 Wireless (i-STAT 1W) using representative cartridges. The studies were based on CLSI guideline EP9-A3.<sup>7</sup> Whole blood samples anticoagulated with lithium heparin were evaluated. Samples were analyzed in duplicate on both systems. A weighted Deming regression analysis was performed using the first replicate result from the i-STAT Alinity versus the mean of the duplicates from the i-STAT 1W.

In the method comparison table, n is the number of specimens, and r is the correlation coefficient.

Test	Units	Comparative Method i-STAT 1W	
pH		n	187
		Slope	0.990

Test	Units	Comparative Method i-STAT 1W	
		r	0.999
		intercept	0.075
		X <sub>min</sub>	6.592
		X <sub>max</sub>	8.189
PO <sub>2</sub>	mmHg	n	192
		Slope	0.986
		r	0.998
		intercept	0.0
		X <sub>min</sub>	9
		X <sub>max</sub>	705
		n	149
		Slope	0.989
PCO <sub>2</sub>	mmHg	r	0.999
		intercept	0.3
		X <sub>min</sub>	5.1
		X <sub>max</sub>	129.8














### FACTORS AFFECTING RESULTS

Factor	Analyte	Effect
Exposing the sample to air	PO <sub>2</sub>	Exposure of the sample to air will cause an increase in PO <sub>2</sub> when values are below 150 mmHg and a decrease in PO <sub>2</sub> when values are above 150 mmHg (approximate PO <sub>2</sub> of room air).
	pH	Exposing the sample to air allows CO <sub>2</sub> to escape which causes PCO <sub>2</sub> to decrease and pH to increase and HCO <sub>3</sub> and TCO <sub>2</sub> to be underestimated.
	PCO <sub>2</sub>	
	HCO <sub>3</sub>	
	TCO <sub>2</sub>	
Venous stasis	pH	Venous stasis (prolonged tourniquet application) and forearm exercise may decrease pH due to localized production of lactic acid.
Hemodilution	pH	Hemodilution of the plasma by more than 20% associated with priming cardiopulmonary bypass pumps, plasma volume expansion or other fluid administration therapies using certain solutions may cause clinically significant error on sodium, chloride, ionized calcium and pH results. These errors are associated with solutions that do not match the ionic characteristics of plasma. To minimize these errors when hemodiluting by more than 20%, use physiologically balanced multi-electrolyte solutions containing low-mobility anions (e.g., gluconate).
Cold temperature	PO <sub>2</sub>	Do not ice samples before testing as PO <sub>2</sub> results may be falsely elevated in cold samples. Do not use a cold cartridge as PO <sub>2</sub> results may be falsely decreased if the cartridge is cold.
Allowing blood to stand (without exposure to air)	pH	pH decreases on standing anaerobically at room temperature at a rate of 0.03 pH units per hour. <sup>1</sup>
	PO <sub>2</sub>	Standing anaerobically at room temperature will decrease PO <sub>2</sub> at a rate of 2–6 mmHg per hour. <sup>1</sup>
	PCO <sub>2</sub>	Standing anaerobically at room temperature will increase PCO <sub>2</sub> by approximately 4 mmHg per hour.
	HCO <sub>3</sub>	Allowing blood to stand (without exposure to air) before testing allows PCO <sub>2</sub> to increase and pH to decrease, which will cause HCO <sub>3</sub> and TCO <sub>2</sub> to be over-estimated, due to metabolic processes.
TCO <sub>2</sub>		
Under fill or partial draw	PCO <sub>2</sub>	The use of partial draw tubes (evacuated tubes that are adjusted to draw less than the tube volume, e.g., a 5 mL tube with enough vacuum to draw only 3 mL) is not recommended due to the potential for decreased PCO <sub>2</sub> , HCO <sub>3</sub> and TCO <sub>2</sub> values. Underfilling blood collection tubes may also cause decreased PCO <sub>2</sub> , HCO <sub>3</sub> and TCO <sub>2</sub> results. Care must be
	HCO <sub>3</sub>	
	TCO <sub>2</sub>	



Factor	Analyte	Effect
		taken to eliminate “bubbling” of the sample with a pipette when filling a cartridge to avoid the loss of CO <sub>2</sub> in the blood.
Method of calculation	sO <sub>2</sub>	Calculated sO <sub>2</sub> values from a measured PO <sub>2</sub> and an assumed oxyhemoglobin dissociation curve may differ significantly from the direct measurement. <sup>3</sup>
Clinical conditions	HCO <sub>3</sub>	Causes of primary metabolic acidosis (decrease calculated HCO <sub>3</sub> ) are ketoacidosis, lactate acidosis (hypoxia), and diarrhea. Causes of primary metabolic alkalosis (increase calculated HCO <sub>3</sub> ) are vomiting and antacid treatment.
Propofol (Diprivan®) or thiopental sodium	PCO <sub>2</sub>	The use of G3+ cartridge is recommended, which is free from clinically significant interference at all relevant therapeutic doses.
PO <sub>2</sub> sensitivity	PCO <sub>2</sub>	In patient samples where the PO <sub>2</sub> is > 100 mmHg above the normal range (80-105 mmHg), an increase in PCO <sub>2</sub> of approximately 1.5 mmHg (with a range of 0.9 to 2.0 mmHg) may be observed for every 100 mmHg increase in PO <sub>2</sub> .  For example, if an oxygenated patient has a measured PO <sub>2</sub> of 200 mmHg, and a normal PO <sub>2</sub> is 100 mmHg, the impact to the PCO <sub>2</sub> result may be increased by approximately 1.5 mmHg.

## KEY TO SYMBOLS

Symbol	Definition/Use
	2 months room temperature storage at 18-30°C
	Use by or expiration date. An expiration date expressed as YYYY-MM-DD means the last day the product can be used.
	Manufacturer's lot number or batch code. The lot number or batch will appear adjacent to this symbol.
	Sufficient for <n> tests
	Authorized representative for Regulatory Affairs in the European Community.
	Temperature limitations. The upper and lower limits for storage are adjacent to upper and lower arms.
	Catalog number, list number, or reference
	Do not reuse.
	Manufacturer
	Consult instructions for use or see System Manual for instructions.
	<i>In vitro</i> diagnostic medical device
	Compliance to the European directive on <i>in vitro</i> diagnostic devices (98/79/EC)
	For prescription use only.

**Additional Information:** To obtain additional product information and technical support, refer to the company website at [www.pointofcare.abbott](http://www.pointofcare.abbott).

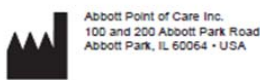
## References

1. Pruden EL, Siggard-Andersen O, Tietz NW. Blood Gases and pH. In: C.A. Burtis and E.R. Ashwood, ed. *Tietz Textbook of Clinical Chemistry*. Second Edition ed. Philadelphia: W.B. Saunders Company; 1994.

2. Tietz NW, Pruden EL, Siggaard-Andersen O. Electrolytes. In: C.A. Burtis and E.R. Ashwood, ed. *Tietz Textbook of Clinical Chemistry*. Second Edition ed. Philadelphia: W.B. Saunders Company; 1994.
3. CLSI. Blood Gas and pH Analysis and Related Measurements; Approved Guideline. *CLSI document C46-A*. 2001.
4. Young DS. *Effects of Drugs on Clinical Laboratory Tests*. 3rd ed. ed. Washington, DC: American Association of Clinical Chemistry; 1990.
5. Painter PC, Cope JY, Smith JL. Reference Ranges, Table 41–20. In: C.A. Burtis and E.R. Ashwood, ed. *Tietz Textbook of Clinical Chemistry*. Second Edition ed. Philadelphia: W.B. Saunders Company; 1994.
6. Statland BE. *Clinical Decision Levels for Lab Tests*. Oradell, NJ: Medical Economic Books; 1987.
7. Clinical and Laboratory Standards Institute (CLSI). Measurement Procedure Comparison and Bias Estimation Using Patient Samples; Approved Guideline—Third Edition. *CLSI document EP09-A3*. 2013.

i-STAT is a trademark of the Abbott Group of companies.

Diprivan is a registered trademark of the AstraZeneca group of companies.



EMERGO EUROPE  
Prinsessegracht 20  
2514 AP The Hague  
The Netherlands



©2019 Abbott Point of Care Inc. All rights reserved. Printed in USA.

