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TwistDx™

Unwind DNA's possibilities

TwistAmp™ Liquid exo

Quick Guide

Part number: INLQEXO

Revision 3

TwistDx™

Abbott House, Vanwall Business Park, Vanwall Road, Maidenhead, SL6 4XE, UK

E info@twistdx.co.uk | twistdx.co.uk

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TwistAmp™ Liquid exo Quick Guide

Please see **instruction and assay design manuals** at twistdx.co.uk for information regarding components and storage, assay design, and detailed use.

Instructions are based on 50 µl reaction volumes; if using a different volume, quantities should be adjusted appropriately.

Primer screen set-up (single-plex)¹

1. Add 2.1 µl of each primer and 0.6 µl of exo probe at 10µM concentration to 0.2 ml PCR tubes.
2. Prepare a pre-master mix (per reaction) in the order below:
2x Reaction Buffer 25 µl
dNTPs² + water³ to 8.2 µl
10x Probe E-mix 5 µl
Vortex and spin briefly.
3. To the pre-master mix, add 2.5 µl 20x Core Reaction Mix⁴ (per reaction) to tube lid. Mix by 10x full inversions and spin briefly.

4. Add 1 µl 50x exo (per reaction) to tube lid. Mix by 10x full inversions and spin briefly. Master mix is now complete⁵. Pipette mix before use.
5. Add 41.7 µl³ master mix to primers and probe prepared in tubes (step 1) and pipette mix.
6. Add 2.5 µl of 280mM MgOAc (supplied) and 1 µl template to tube lids³. DNA and MgOAc should be kept separate in the tube lid prior to spin-down. Spin in MgOAc/template and mix well (6x inversions) to start reaction. Spin briefly.

Warning: RPA reactions start as soon as MgOAc is added.

7. Place reactions in a fluorometer and start run: 37-42°C, 20 minutes. For low template copies, remove strip after 4 mins, mix by 6x full inversions and spin briefly, replace in fluorometer.

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Template screen set-up (single-plex)¹

1. Prepare a primer and probe pre-master mix (per reaction) in the order below:
2x Reaction Buffer 25 µl
dNTPs² + water³ to 8.2 µl
10x Probe E-mix 5 µl
Primer A (10µM) 2.1 µl
Primer B (10µM) 2.1 µl
Probe (10µM) 0.6 µl
Vortex and spin briefly.
2. Add 2.5 µl 20x Core Reaction Mix⁴ (per reaction) to tube lid. Mix by 10x full inversions and spin briefly.
3. Add 1 µl 50x Exo (per reaction) to tube lid. Mix by 10x full inversions and spin briefly. Master mix is now complete⁵. Pipette mix before use.
4. Add 46.5 µl³ master mix to 0.2 ml PCR tubes.
5. Add 2.5 µl of 280mM MgOAc and 1µl template to tube lid³. DNA and MgOAc should be kept separate in the tube lid prior to spin-down. Spin in MgOAc/template and mix well (6x inversions) to start reaction. Spin briefly.

Warning: RPA reactions start as soon as MgOAc is added.

6. Place reactions in fluorometer and start run: 37-42°C, 20 minutes. For low template copies, remove strip after 4 mins, mix by 6x full inversions and spin briefly, replace in fluorometer.

¹ See manual for multiplexing.

² Suggested final concentration of 1.8mM (total) dNTPs. Optimisation is recommended.

³ Volumes should be adjusted if adding more/less template and/or MgOAc.

⁴ Warm to room temperature and pipette mix slowly to ensure homogeneity.

⁵ Master mix may appear cloudy, this is normal.

TwistDX
Liquid exo QG

Size:
8.268 in x 5.827 in



PMS 185 C
Red



Black



PMS 7541 C
Gray1 - 10%

Part Number: INLQEXO
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